



UNIVERSITÀ
DEGLI STUDI
FIRENZE

FLORE

Repository istituzionale dell'Università degli Studi di Firenze

3-ALFA-TROPANYL 2-(4-CI-PHENOXY)BUTIRATE (SM21): A REVIEW OF THE PHARMACOLOGICAL PROFILE OF A NOVEL ENHANCER OF

Questa è la Versione finale referata (Post print/Accepted manuscript) della seguente pubblicazione:

Original Citation:

3-ALFA-TROPANYL 2-(4-CI-PHENOXY)BUTIRATE (SM21): A REVIEW OF THE PHARMACOLOGICAL PROFILE OF A NOVEL ENHANCER OF CHOLINERGIC TRANSMISSION / C. GHELARDINI; N. GALEOTTI; F. GUALTIERI; S. SCAPECCHI; A. BARTOLINI. - In: CNS DRUG REVIEWS. - ISSN 1080-563X. - STAMPA. - 3:(1997), pp. 346-362. [10.1002/(SICI)1098-2299(199705)41:11::AID-DDR1>3.0.CO;2-M]

Availability:

This version is available at: 2158/1207 since:

Published version:

DOI: 10.1002/(SICI)1098-2299(199705)41:1<1::AID-DDR1>3.0.CO;2-M

Terms of use:

Open Access

La pubblicazione è resa disponibile sotto le norme e i termini della licenza di deposito, secondo quanto stabilito dalla Policy per l'accesso aperto dell'Università degli Studi di Firenze (<https://www.sba.unifi.it/upload/policy-oa-2016-1.pdf>)

Publisher copyright claim:

(Article begins on next page)

3- α -tropanyl 2-(4-Cl-phenoxy)butyrate (SM 21): A Review of the Pharmacological Profile of a Novel Enhancer of Cholinergic Transmission

C. Ghelardini, N. Galeotti, F. Gualtieri,*
S. Scapecchi,* and A. Bartolini

*Department of Pharmacology and *Department of Pharmaceutical Sciences,
University of Florence, Florence, Italy*

Key Words: Acetylcholine release—Analgesia—Antinociception—Cholinergic system—
Learning and memory—Pain—SM 21.

INTRODUCTION

Atropine-like preparations were used by ancient Romans to relieve pain; Pliny the elder, in his *Historia Naturalis*, reported that the juice of *Mandragora officinarum* or *Hyoscyamus niger* was administered to patients before surgery to produce analgesia. Much more recently Ghelardini et al. (31) confirmed the paradoxical effect of atropine by reporting that, at very low doses, this compound induces central antinociception in rodents through an enhancement of cholinergic transmission. It is interesting to note that this antinociceptive activity, unlike that produced by direct muscarinic agonists and cholinesterase inhibitors, was not accompanied by typical cholinergic symptomatology symptoms (tremors, sialorrhea, diarrhea, rhinorrhea, lacrimation, etc.). Soon after, it was discovered that the R-(+)-enantiomer of atropine, R-(+)-hyoscyamine, was responsible for the antinociceptive activity of the racemate, while the S-(–)-enantiomer, S-(–)-hyoscyamine, was devoid of any antinociceptive action (33). R-(+)-hyoscyamine, in the same range of analgesic doses, was also able to prevent amnesia induced by antimuscarinic drugs (41). An investigation of the antinociceptive and anti-amnesic effect of atropine, using microdialysis techniques has demonstrated that R-(+)-hyoscyamine, at cholinomimetic doses, produced an increase in the acetylcholine (ACh) release from the rat cerebral cortex *in vivo*, indicating that this compound has a presynaptic mechanism of action (41).

Based on these observations, a program to modify the chemical structure of atropine was started, aimed at developing cholinergic amplifiers endowed with more in-

Address correspondence and reprint requests to Prof. A. Bartolini, Department of Pharmacology, University of Florence, viale G. B. Morgagni 65, I-50134 Florence, Italy. Fax: 39-55-4361613.

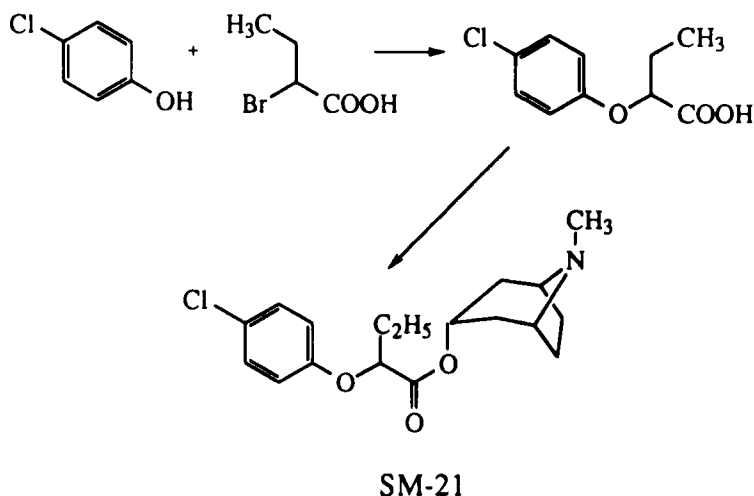


Fig. 1. Chemical structure and synthesis of SM 21.

tensive antinociceptive and anti-amnesic activity than atropine, but with the same lack of cholinergic side effects as atropine. These compounds would be potentially useful as analgesics and/or in the treatment of pathological conditions, such as Alzheimer's disease, characterized by cholinergic deficit. Of the many compounds synthesized and studied, SM 21 (3- α -tropanyl 2-[4-Cl-phenoxy]butyrate) (48) (Fig. 1) showed the best pharmacological profile.

CHEMISTRY

Modification of the atropine structure by substituting the phenyl ring or the aminoalcohol moiety provided some potent compounds whose efficacy, compared with morphine, remained as low as that of atropine (48). Better results were obtained in the series of 2-phenylpropionic acid esters, which were synthesized to deal with the chemical instability of tropic acid. In this class the potency was much lower than that of atropine, but efficacy was definitely improved (48).

To restore high affinity, it was thought that the possibility of a hydrogen bond, present in atropine, should be reintroduced into the molecule. We synthesized several esters of substituted 2-(phenoxy)propionic acids. Further modifications of the molecule showed that 2-(phenoxy)butyric acid gave better results than the corresponding 2-(phenoxy)propionic acid and that the 3- α -tropanol was the best choice for the aminoalcohol moiety. In this class, SM 21 was selected as the most interesting compound (47). Its chemical structure and synthesis are illustrated in Fig. 1. Isosteric substitution of the oxygen atom with S, NH, NCH₃, or CH₂ was also performed (47).

Chemical modifications have led to compounds similar to SM 21, such as PG 9 (6), ET 142 (38), and SM 32 (40), with a pharmacological profile quite similar to that of

SM 21. Other chemical modifications, though informative regarding structure-activity relationships in the series, produced less interesting compounds (64).

SM 21 possesses a stereogenic center and, as a consequence, is normally obtained as a racemic mixture of two enantiomers. To develop the compound further, it was necessary to study the properties of the single enantiomers; therefore, we addressed the problem of obtaining the two enantiomers with acceptable optical purity (76).

SM 21 enantiomers show a certain enantioselectivity in pharmacological activity even if both stereoisomers are active (39) unlike what happens for atropine enantiomers, where only R-(+)-hyoscyamine shows analgesic and antiamnesic activity (41). In any case, the most potent and efficacious enantiomer is R-(+)-SM 21, which shares the same absolute configuration of R-(+)-hyoscyamine.

CENTRAL PHARMACOLOGICAL PROFILE

In Vivo Studies

Antinociceptive Properties

SM 21 induced antinociception in mice, rats, and guinea pigs. Antinociception was elicited regardless of which noxious stimulus was used: thermal (hot-plate and tail flick tests), chemical (abdominal constriction test), or mechanical (paw pressure test), performed according to O'Callaghan and Holtzman (69), D'Amour and Smith (17), Koster et al. (56), and Leighton et al. (59), respectively.

SM 21 produced a dose-dependent increase in the pain threshold in mice after systemic (subcutaneous [s.c.], intraperitoneal [i.p.], oral [p.o.], intravenous [i.v.]) injection, as illustrated by the hot-plate (Fig. 2) and abdominal constriction tests (Fig. 3a). SM 21 reached its maximum antinociceptive effect 15 min after administration and then slowly diminished (Fig. 2b,d). SM 21 produced an increase in the pain threshold not only in mice but also in rats, in the paw pressure (Fig. 3b), and tail flick (39) tests and in guinea pigs, in the paw pressure test (39), with a pharmacological profile similar to that exerted in mice.

SM 21 is endowed with central antinociceptive activity. It was, in fact, possible to reach the same intensity of analgesia by injecting directly into the cerebral ventricles (49) doses (5 to 20 $\mu\text{g}/\text{mouse}$) of SM 21 that were fifty times lower than those needed parenterally (Fig. 2c). That the antinociception depends on a retrodiffusion of the drug from the cerebral ventricles to the periphery can thus be ruled out.

Both enantiomers of SM 21, R-(+)-SM 21, and S-(–)-SM 21 induced antinociception in the mouse hot-plate and abdominal constriction tests in a dose-dependent manner; R-(+)-SM 21 was slightly more effective than S-(–)-SM 21 (39).

SM 21 showed good antinociceptive efficacy in comparison with that produced by R-(+)-hyoscyamine and some well known analgesic drugs such as morphine, diphenhydramine, and clomipramine. As a matter of fact, by comparing the areas under the curve of the above-mentioned compounds, tested at the highest doses that do not im-

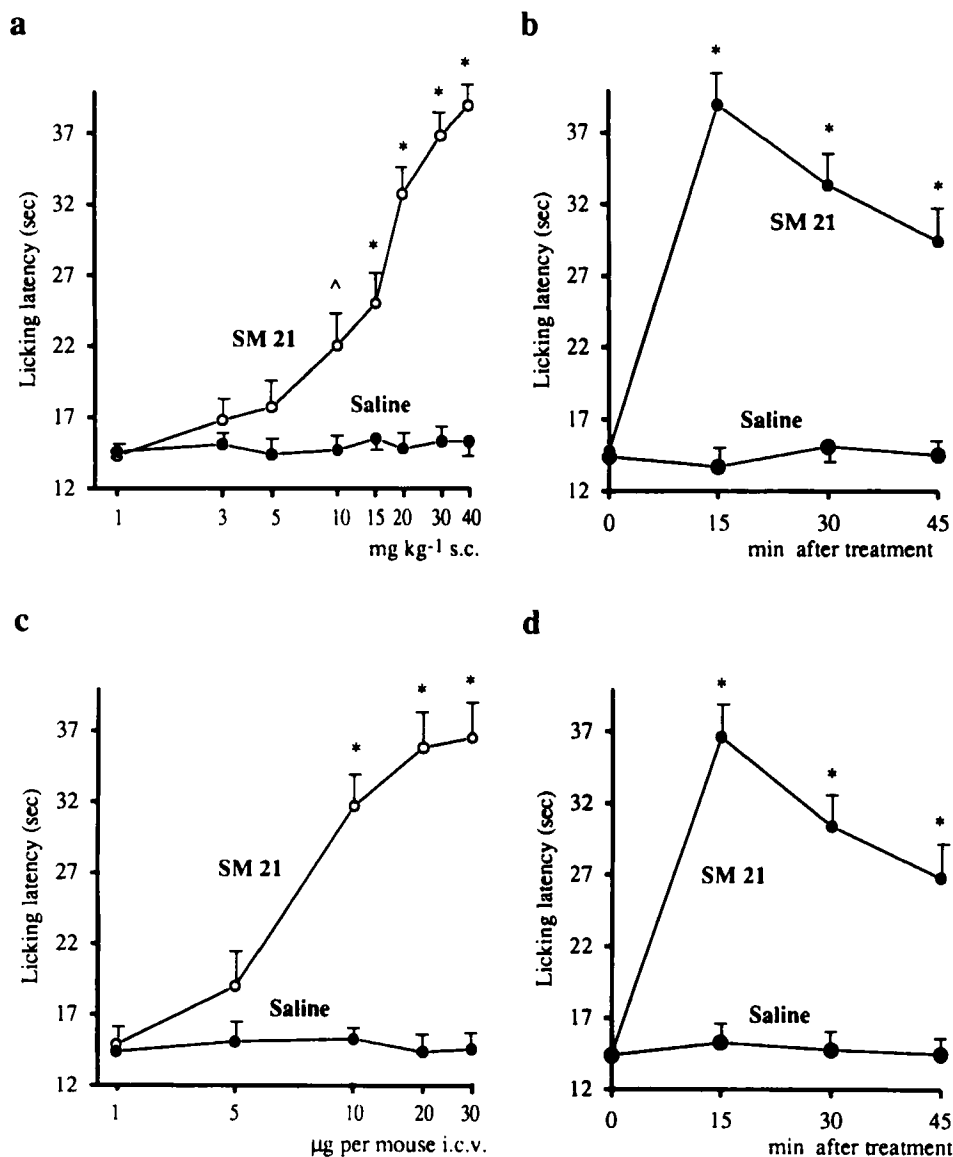


Fig. 2. Dose-response curves of SM 21 i.p. (a) and i.c.v. (c) injected in the mouse hot-plate test. The SM 21 time course of 40 mg/kg i.p. is reported in (b) and 30 µg per mouse i.c.v. in (d) from the same test. Vertical lines give S.E.M. Each point is the mean of at least 10 mice. [^] $P < 0.05$; * $P < 0.01$ in comparison with saline controls. In (a) and (c) SM 21 was administered 15 min before the test.

pair mouse normal behavior, SM 21 was as effective as morphine, and more effective than R-(+)-hyoscyamine, diphenhydramine and clomipramine (39).

SM 21, at doses lower than 1 mg/kg, was able to reduce the number of abdominal constrictions induced by intraperitoneal injection of a 0.3% acetic acid solution and to

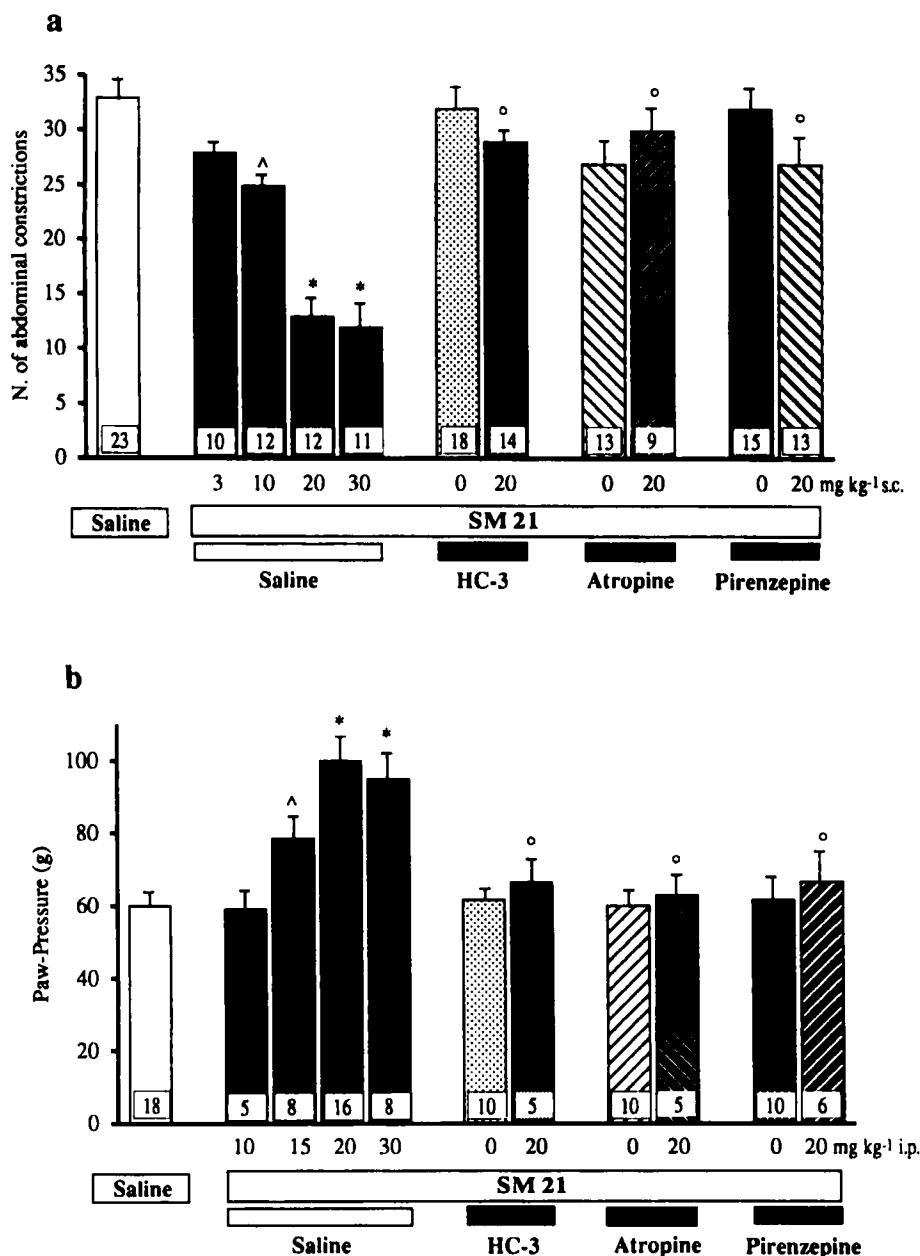


Fig. 3. (a) Antinociceptive effect of SM 21 and antagonism of hemicholinium-3 (HC-3) (1 μ g per mouse i.c.v.), atropine (5 mg/kg i.p.) and pirenzepine (0.1 μ g per mouse i.c.v.) on the enhancement of pain threshold induced by SM 21 (20 mg/kg s.c.) in the mouse abdominal constriction test induced by 0.6% acetic acid (a) and in the rat paw-pressure tests (b) HC-3, atropine and pirenzepine were injected respectively 5 h, 15 min, and 10 min before testing. In the abdominal constriction test the nociceptive responses were recorded 15 min after SM 21 administration. Vertical lines show S.E.M. $^{\wedge}P < 0.05$; $^*P < 0.01$ in comparison with saline controls. $^{\circ}P < 0.01$ in comparison with SM 21 (20 mg/kg s.c.). Numbers inside the columns indicate the number of mice or rats.

reverse the hyperalgesia induced by morphine withdrawal (data not shown). SM 21 antinociception is not due to an anti-inflammatory action. SM 21, at concentrations up to 10^{-4} M, did not inhibit inducible COX activity in comparison with indomethacin (IC_{50} : 23×10^{-6} M) and, at analgesic doses, failed to suppress paw edema in response to carrageenan administration (Table 1).

Antiamnesic Activity

SM 21 ameliorated cognitive processes in mice and rats. This compound was able to prevent amnesia induced by treatment with drugs such as scopolamine (Fig. 4), dicyclomine (Fig. 4), diazepam (27), and AF64A (26), or exposure to hypoxic environment (67) in the passive avoidance test. The antiamnesic effect of SM 21 was dose-dependent and the first active dose was lower than that able to enhance the pain threshold. A complete prevention of amnesia was, in fact, obtained at a dose (10 mg/kg) that was weakly analgesic only in the hot-plate test. The time-course of the antiamnesic activity of SM 21 was equal to that observed for the antinociceptive action, reaching its maximum effect between 15 and 30 min after injection. Therefore, in the passive avoidance experiments SM 21 was administered 20 min before the training session.

In the passive avoidance test an improvement in cognition of animals that have no memory impairment is difficult to demonstrate. SM 21, as well as well-known nootropic drugs, such as piracetam and aniracetam, or cholinomimetics, such as physostigmine and oxotremorine, do not show any memory facilitation in unamnesic animals (45,16).

A procognitive activity of SM 21 was unmasked by using a social learning test, performed according to Mondadori et al. (68), but in which adults rats with unimpaired memory were used. SM 21, as well as piracetam, exerted beneficial effects on the cognitive performance by prolonging the time spent by rats to delete mnemonic information (25).

Subacute Treatment

SM 21 induced tolerance after repeated administration. SM 21, injected twice daily for two weeks at doses at which it demonstrates a full antiamnesic and antinociceptive

TABLE 1. *Effect of SM 21 on carrageenan-induced paw edema in rats*

Pretreatment	Treatment	Dose, mg/kg i.p.	Paw volume ml \pm S.E.M.
Saline	Saline		1.37 \pm 0.08
Carrageenan	Saline		2.21 \pm 0.09
Carrageenan	SM 21	20	2.27 \pm 0.06
Carrageenan	SM 21	30	2.19 \pm 0.05
Carrageenan	Indomethacin	1	1.45 \pm 0.07*

Indomethacin was used as positive control; $n = 5$ rats per group. * $P < 0.05$ in comparison with carrageenan-saline controls. SM 21 was injected 15 min before the test.

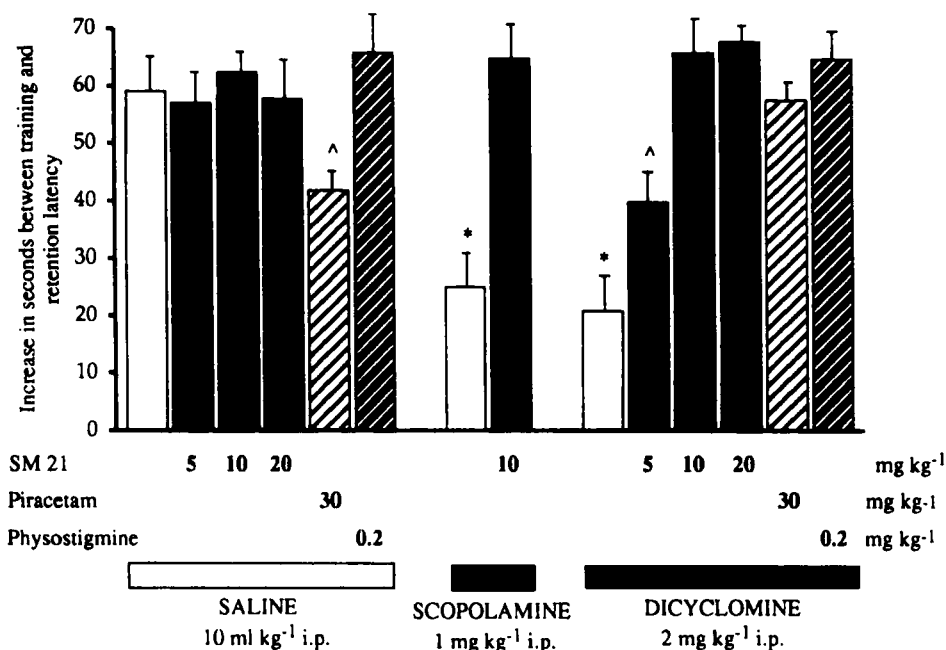


Fig. 4. Effect of i.p. SM 21, piracetam, and physostigmine on dicyclomine-induced amnesia in mouse passive avoidance test and, under the same experimental conditions, effect of SM 21 on scopolamine amnesia. Punishment consists of a fall into cold water (10°C). SM 21, piracetam, and physostigmine were injected 20 min before the training session. Scopolamine and dicyclomine were injected immediately after the training session. ^ $P < 0.05$; * $P < 0.01$ in comparison with saline controls. Each column represents the mean of at least 25 mice.

activity (10 and 30 mg/kg i.p., respectively), produced a complete loss of both behavioral effects. Following the same administration schedule, however, other analgesic drugs, such as morphine, oxotremorine, and baclofen, develop tolerance toward their analgesic effect (61,62). Subacute treatment with SM 21 (30 mg/kg i.p.) did not produce loss of body weight or the symptomatology typical of the withdrawal syndrome.

Effect of SM 21 on Animal Behavior

The maximum antinociceptive effect of SM 21 was obtained at 40 mg/kg s.c. without producing any visible modification in mouse or rat gross behavior. At the same dose, SM 21-treated mice showed a complete integrity of motor coordination on the rotarod test, tested according to Kuribara et al. (57) (Table 2). Under these experimental conditions, SM 21 was compared with equiactive doses of oxotremorine and physostigmine (Table 2). The muscarinic agonist and the inhibitor of cholinesterase both produced a statistically significant reduction in endurance time on the rotating rod. Normal spontaneous motility, evaluated by the Animex apparatus (data not shown), as well as exploratory behavior, revealed by the hole-board test (39), were

TABLE 2. *Effect of SM 21, oxotremorine, and physostigmine in the rotarod test*

Treatment s.c.	Endurance time on rotarod (s)			
	Before treatment	After treatment		
		15 min	30 min	45 min
Saline	98.5 ± 5.1 (16)	92.7 ± 6.2 (16)	103.7 ± 5.0 (16)	97.4 ± 7.2 (16)
SM 21, 40mg/kg	103.2 ± 5.6 (10)	97.5 ± 6.2 (10)	99.6 ± 5.4 (10)	100.2 ± 4.3 (10)
Oxotremorine, 40 µg/kg	106.2 ± 8.2 (11)	76.5 ± 7.3* (11)	63.6 ± 9.6* (11)	64.4 ± 8.7* (11)
Physostigmine, 200 µg/kg	93.4 ± 5.7 (9)	61.4 ± 6.8* (9)	54.5 ± 8.1* (9)	52.3 ± 8.8* (9)

* $P < 0.05$ in comparison with saline controls. The number of mice is shown in parentheses.

also observed after subcutaneous administration of SM 21 at 40 mg/kg s.c. and intracerebroventricular (i.c.v.) administration of 30 µg/mouse. Impaired motor coordination and spontaneous motility were revealed in mice starting at 100 mg/kg s.c. The LD₅₀ was at 400 mg/kg s.c., corresponding to 883 µmol/kg.

PERIPHERAL PHARMACOLOGICAL PROFILE

Effect on Smooth Muscle

Effect on Intestinal Motility

SM 21, administered at analgesic and anti-amnesic doses, did not modify transit in the intestinal tract of the mouse, performed according to Reynell and Spray (74) (data not shown). In contrast, other analgesic drugs, such as morphine, significantly retarded gastrointestinal propulsion, the cholinesterase inhibitor neostigmine accelerated net propulsion (80). The lack of effect of SM 21 on intestinal motility indicates that this compound, with the same analgesic activity, has advantage over opioid analgesics, which produce constipation, or classical cholinomimetics, which produce diarrhea.

Effect on Isolated Guinea Pig Ileum

SM 21 added to the organ bath at concentrations ranging from 1 pM to 1 nM potentiated the contractions evoked by both nicotine (4 µM) and electrical stimulation at 0.1 Hz, 0.5 ms, voltage double threshold, performed according to Paton and Vizi (71) (Fig. 5). The effect was larger (area under the curve ratio) on the contractions induced by nicotine than those induced by electrical stimulation. The potentiation was no longer observed when the concentration of SM 21 in the medium was raised to 10 nM.

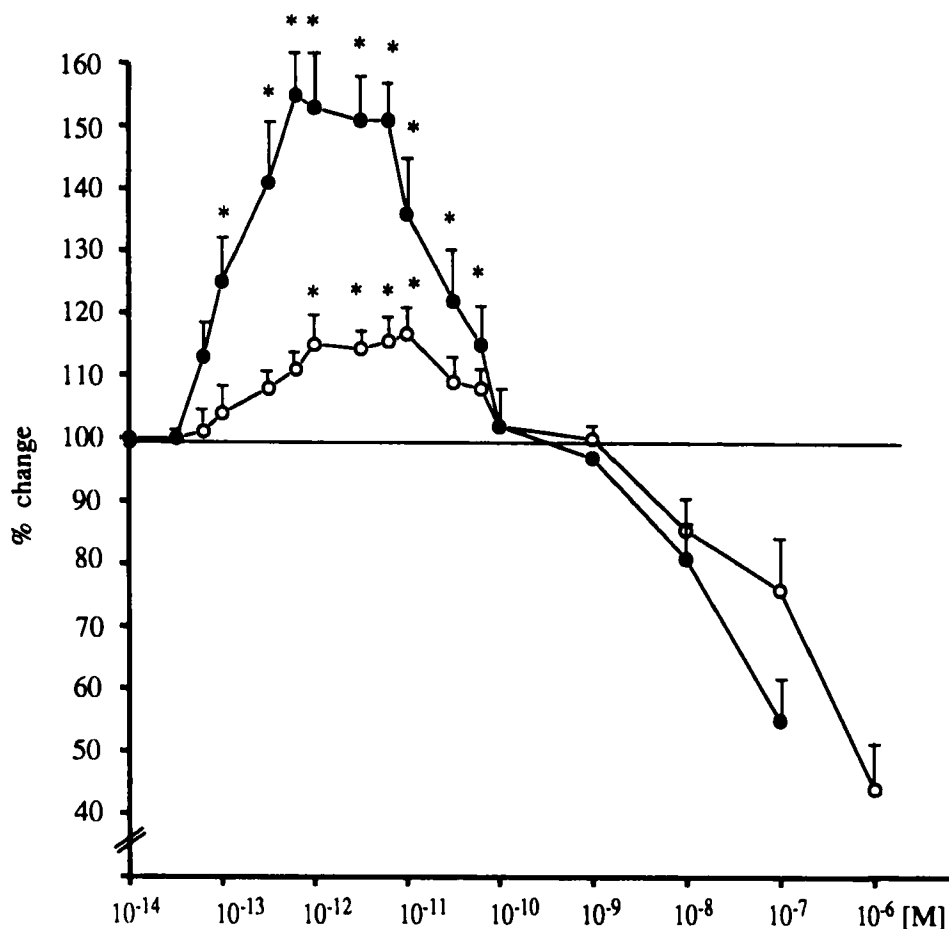


Fig. 5. Dose-response curves of SM 21 on nicotine ($4 \mu\text{M}$; closed symbols) and electrically (0.1 Hz ; 0.5 ms ; double threshold voltage; open symbol evoked contractions) of guinea pig ileum myenteric plexus longitudinal muscle strip expressed as percentage variation of contractions. Each point represents the mean of at least 6 experiments and vertical lines give S.E.M. * $P < 0.05$ calculated in the range between 0.1 pM and 0.1 nM .

SM 21 began to inhibit both types of evoked contractions at $1 \mu\text{M}$. Nicotine-evoked ileum contractions were about four times greater than those electrically evoked (Fig. 5). This is probably due to the simultaneous activation, during electrically-evoked contractions, of both intramural cholinergic and sympathetic fibers, whereas during nicotine-evoked contractions only cholinergic neurons are likely to be activated. Noradrenaline released during electrical stimulation could be responsible for limiting the effect of the ACh released by low doses of SM 21. The higher amplification by SM 21 of the nicotine-evoked contractions of guinea pig ileum as compared

with those elicited by electrical stimulation depended on the inhibitory control exerted by norepinephrine, which is released only during electrical stimulation (31).

Effect on Striated Muscle

Rat Phrenic Nerve-Hemidiaphragm Preparation

SM 21 (1 μ M to 1 mM) potentiated the hemidiaphragm contractions evoked by electrical stimulation of the left phrenic nerve, performed according to the method described by Bülbring (10) and modified by Wessler and Kilbinger (83), and did not modify the contractions evoked through direct stimulation of the diaphragm muscle (data not shown). At concentrations lower than 1 μ M, SM 21 was always inactive. A potentiation of hemidiaphragm contractions is exerted by numerous muscarinic antagonists, such as atropine, pirenzepine, dicyclomine, and glycopyrrolate (73,84) by blocking the muscarinic autoreceptors. SM 21 may, therefore, exert its effect on the phrenic nerve by antagonizing muscarinic receptor subtypes. One must consider, however, that inhibitors of cholinesterase can also amplify hemidiaphragm contractions (1). Since SM 21 is endowed with very low anticholinesterase activity ($IC_{50} = 110 \mu$ M), it may be possible that its action underlies antimuscarinic and/or anticholinesterase activity. The lack of inhibition of electrical stimulation of hemidiaphragm contractions rules out the possibility that SM 21 acts as a local anesthetic. In fact, local anesthetics such as lidocaine and procaine inhibit the electrically stimulated contractions of the same preparation up to complete abolishment in a dose-dependent manner (1).

MECHANISM OF ACTION

SM 21 antinociception was found to be dependent on cholinergic activation, since this analgesia is antagonized by the muscarinic antagonist atropine (Fig. 3a,b), the M_1 -antagonist pirenzepine (Fig. 3a,b), the ACh depletor HC-3 (Fig. 3a,b), and by lesion of the nucleus basalis magnocellularis (NBM) (6), which is the primary source of ACh for the cerebral cortex (75). Moreover, the antagonism exerted by intracerebroventricular injection of HC-3 and pirenzepine in mice and NBM lesions in rats on SM 21-induced antinociception confirms that the site of action of SM 21 is centrally located.

A presynaptic mechanism facilitating cholinergic transmission is involved in SM 21 activity as revealed by microdialysis studies performed according to Giovannini et al. (43). SM 21 increased ACh release from rat cerebral cortex, which peaked from 45 to 60 min after administration and returned to basal values within 120 min (Fig. 6). This effect was sensitive to tetrodotoxin (Fig. 6). The SM 21-induced increase in ACh release occurred at the same range of doses (10 to 20 mg/kg i.p.) at which SM 21 exerted its antinociceptive and anti-amnesic activities. The greater latency required to reach the maximum amplification of ACh release compared to that

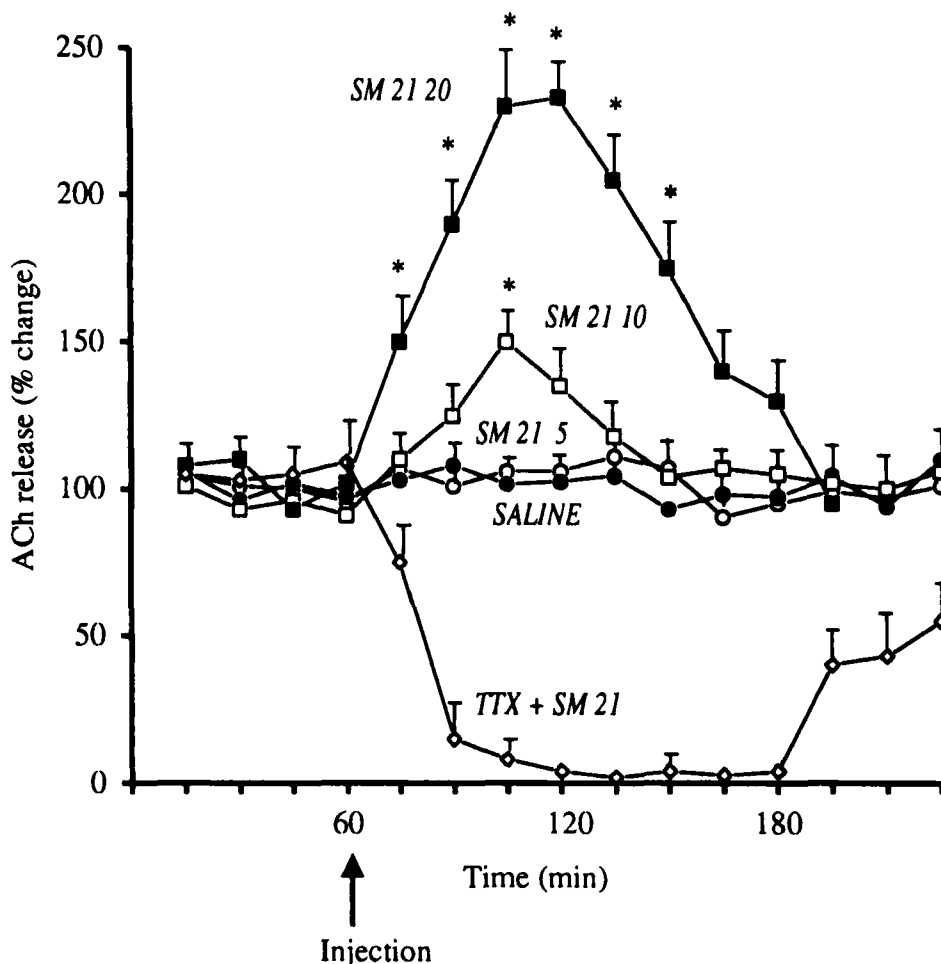


Fig. 6. Dose-response curves of SM 21 on ACh release from parietal cortex and antagonism of SM 21 (20 mg/kg i.p.) by TTX (0.5 μ M). All values are expressed as changes over basal output. SM 21 was administered at 60 min as shown by the arrow. Vertical lines give S.E.M. Each point represents the mean of at least 5 independent experiments. Doses of SM 21 are expressed as mg/kg i.p. Significant differences were evaluated by comparing the percentage variation vs. the mean \pm S.E.M. of all pre-drug determinations. * $P < 0.05$ in comparison with controls.

required to be active could be ascribed to the time taken by ACh to diffuse from the synaptic cleft to the microdialysis tube.

The hypothesis of a presynaptic cholinergic mechanism for SM 21 is confirmed by: 1) the SM 21-induced amplification of electrically and chemically evoked contractions of guinea pig ileum myenteric plexus longitudinal muscle strips (Fig. 5) without modifying its basal tone; 2) the antagonism of SM 21-induced antinociception by the ACh depletor HC-3. A postsynaptic mechanism of action for SM 21 can be ruled out since, as reported by Bartolini et al. (3,5), HC-3 was not able to antagonize antinociception

TABLE 3. Affinity profiles of SM 21, R-(+)-hyoscyamine, and AFDX-116 at M_1 – M_4 muscarinic receptors and binding affinities of SM 21 and AFDX-116 for m_1 – m_4 muscarinic receptor subtypes expressed in Chinese hamster ovary cells (CHO-K1)

	pA ₂ values			
	M ₁ rabbit vas deferens	M ₂ rat left atrium	M ₃ rat ileum	M ₄ -putative guinea pig uterus
SM 21	5.97 ± 0.11*	6.63 ± 0.10*	6.35 ± 0.04*	6.26 ± 0.05*
R-(+)-hyoscyamine	7.05 ± 0.05 ^a	7.25 ± 0.04 ^a	6.88 ± 0.05 ^a	9.56 ± 0.01 ^a
AFDX 116	6.84 ± 0.14 ^b	7.12 ± 0.11 ^b	6.34 ± 0.13 ^c	6.70 ± 0.06

	pK _i values			
	m ₁	m ₂	m ₃	m ₄
SM 21	6.90 ± 0.16	6.28 ± 0.12	6.62 ± 0.10	6.53 ± 0.05
AFDX 116	6.84 ± 0.14 ^b	7.12 ± 0.11 ^b	6.34 ± 0.13 ^b	6.70 ± 0.06

Each value represents the mean ± S.E.M.; * pK_B values were obtained with SM 21 1 μM. From ref. ^a34, ^b23, ^c24.

induced by agonists of postsynaptic muscarinic receptors such as oxotremorine, McN-A-343, and AF-102B; and 3) SM 21 did not elicit the typical cholinergic symptoms (tremors, sialorrhea, diarrhea, rhinorrhea, lacrimation, etc.) produced by injection of direct postsynaptic muscarinic agonists (9). It is also to be noted that there is a wide gap between the low concentrations at which SM 21 is thought to inhibit the presynaptic muscarinic receptors (Fig. 5) and the high concentrations that are needed to block the postsynaptic muscarinic receptors (Table 3).

It is well known that activation of the nicotinic system induces antinociception. SM-21, even if it increases extracellular levels of ACh, produces an enhancement of the pain threshold that is not prevented by mecamylamine, excluding a mechanism of action involving the interaction with nicotinic receptors (data not shown). This hypothesis is also supported by the fact the antimuscarinic drugs, at doses able to antagonize muscarinic antinociception, do not prevent nicotinic antinociception (33).

It has long been known that activation of the cholinergic system induces antinociception (72,30,52,51,13,50,60), as well as a facilitation of cognitive processes (16). It is plausible, therefore, that enhancement of extracellular levels of ACh can be considered responsible for the antinociceptive effect of SM 21. Moreover, the SM 21-induced amplification of endogenous ACh release may counteract the amnesic effect produced by the antimuscarinic drugs scopolamine and dicyclomine.

ACh release can be increased by blocking M₂/M₄ muscarinic autoreceptors (58,81,65,79). The affinity profile of SM 21 vs. M₁ (rabbit vas deferens, according to Eltze [23] and modified by Dei et al. [18]), M₂ (guinea pig atrium, according to Eltze et al. [22] and modified by Dei et al. [18]), M₃ (guinea pig ileum, according to Eltze and Figala [24]), and putative M₄ receptors (prepuberal guinea pig uterus, according to Dörje et al. [20]) shows low M₄/M₁ (1.9 times) and M₂/M₁ (4.6 times) selectivity ratios as reported in Table 3. In this study, SM 21 selectivity was compared with that of

the selective M_4 antagonist R-(+)-hyoscyamine (34) and the selective M_2 antagonist AFDX-116* (42). It is possible that a selectivity ratio of 4.6, even if small, may be high enough to enhance the pain threshold and to reverse amnesia as a consequence of ACh release. The M_2 muscarinic antagonists, AFDX-116 (42), methoctramine (66), and AQRA-741 (19), which are endowed, like SM 21, with cholinergic presynaptic antinociceptive (4,32,46) and anti-amnesic (2) properties, and which are able to increase ACh release (58,81), have an M_2/M_1 selectivity ratio comparable to that of SM 21. However, binding studies performed on the m_1 to m_4 human muscarinic receptor subtypes expressed in CHO cells (21,12) did not confirm the results obtained by functional studies, as shown in Table 3. Other mechanisms able to potentiate the endogenous cholinergic system may be involved in the antinociceptive and anti-amnesic effect induced by SM 21.

It has been demonstrated that D_2 dopaminergic (44,82,78,54), A_1 adenosinergic (55,11), H_3 histaminergic (14), 5-HT $_4$ serotonergic heteroreceptors (15), and 5-HT $_{1A}$ receptors (8), increase ACh release. However, the above-mentioned receptors are not involved in an SM 21 mechanism of action. In fact, SM 21 is able to interact with D_2 , H_3 , 5-HT $_4$, and 5-HT $_{1A}$ only at concentrations higher than 10^{-6} M, as revealed by binding studies (data not shown). These results are supported by the fact that quinpirole (D_2 agonist), N^6 -cyclopentyladenosine (A_1 agonist), R-(α)-methylhistamine (H_3 agonist), GR-48125 (5-HT $_4$ antagonist), and NAN 190 (5-HT $_{1A}$ antagonist), at doses able to prevent the antinociception induced respectively by haloperidol (33), caffeine (37), thioperamide (63), BIMU 1 and BIMU 8 (36), and 5-HT $_{1A}$ agonists (35,29), failed to prevent SM 21 antinociception (39).

Neurotransmitter systems other than the cholinergic are not involved in SM 21 antinociception. This compound interacts with the following receptor subtypes: α_1 -, α_2 -, β_1 -, β_2 -adrenoceptors, D_1 , GABA $_A$, GABA $_B$, H_1 , NK $_1$, δ -, κ -, μ -opioid, 5-HT $_{1D}$, 5-HT $_2$, 5-HT $_3$, and K^+ channels: ATP-sensitive K^+ channel, voltage-dependent K^+ channel, Ca^{2+} -activated K^+ channel only at concentrations higher than 10^{-6} M (data not shown). The lack of prevention of SM 21 antinociception by the opioid antagonist naloxone, the GABA $_B$ antagonist CGP-35348 and the biogenic amine depletor reserpine (39) is in agreement with the binding data. Pertussis toxin (PTX) pretreatment was able to prevent opioid (70), catecholaminergic, GABAergic (53), histaminergic (28), and purinergic (77) analgesia, but not muscarinic antinociception (28). Since SM 21 antinociception was not prevented by pretreatment with pertussis toxin (7), the hypothesis of a cholinergic mechanism underlying the SM 21 mechanism of action is further supported.

SUMMARY

SM 21 is a 2-phenylpropionic acid ester, structurally related to atropine, that produces a central antinociceptive and anti-amnesic effect in mice and rats. These activities are exerted without impairing motor coordination and without producing typical cholinergic symptomatology. SM 21 is also able to amplify the evoked contractions

* Chemical name of AFDX-116 is: 11,2-(diethylamino)methyl-1-piperidinyl acetyl-5,11-dihydro-6H-pyrido 2,3b 1,4-benzodiazepine-6-one.

of smooth and striated muscle. A potentiation of endogenous cholinergic activity, by enhancing ACh extracellular levels, can be considered responsible for the action of SM 21 on both the central and peripheral nervous system. However, at this point the exact mechanism by which ACh levels are increased is not entirely elucidated.

REFERENCES

1. Abbs CT, Wall AH. The effect of local anaesthetics on neuromuscular transmission in the rat phrenic nerve-diaphragm preparation. In: Ganguly JK, ed. *Comparative effect of local anaesthetics*. London: Pergamon Press, 1981;22–31.
2. Aura J, Servö, Riekkinen P Jr. Methocitramine moderately improves memory but pirenzepine disrupts performance in delayed non-matching to position test. *Eur J Pharmacol* 1977;333:1209–1234.
3. Bartolini A, Galli A, Ghelardini C, et al. Antinociception induced by systemic administration of local anaesthetics depends on a cholinergic mechanism. *Br J Pharmacol* 1987;92:711–721.
4. Bartolini A, Ghelardini C, Gualtieri F, et al. I.c.v. AFDX 116 induces analgesia only when administered at very low doses. *Trends Pharmacol Sci Suppl* IV 1989:99.
5. Bartolini A, Ghelardini C, Fantetti L, Malcangio M, Malmberg-Aiello P, Giotti A. Role of muscarinic receptor subtypes in central antinociception. *Br J Pharmacol* 1992;105:77–82.
6. Bartolini A, Ghelardini C, Giovannini MG, et al. Modulators of ACh release as potent cognition enhancers and analgesics: Pharmacodynamic studies. *XXVII Symposium of the Italian Pharmacological Society*. Turin, Italy;1994:29.
7. Bartolini A, Ghelardini C, Galeotti N, Beneforti E, Zoppi M. Pharmacology of experimental analgesic, drugs. *Pain, Rheumatic Diseases and Quality of Life*. Florence, Italy;1997:48.
8. Bianchi C, Siniscalchi A, Beani L. 5-HT_{1A} agonists increase and 5-HT₃ agonists decrease acetylcholine efflux from the cerebral cortex of freely-moving guinea pigs. *Br J Pharmacol* 1990;101:448–452.
9. Brown JH, Taylor P. Muscarinic receptor agonists and antagonists. In: Hardman JG, Limbird LE, eds. *Goodman & Gilman's The Pharmacological Basis of Therapeutics*. New York: McGraw-Hill, 1996;141–160.
10. Bülbring E. Observations on the isolated phrenic nerve diaphragm preparation on the rat. *Br J Pharmacol* 1946;1:38–61.
11. Carter AD, O'Connor WT, Carter MJ, Ungerstedt U. Caffeine enhances acetylcholine release in the hippocampus *in vivo* by a selective interaction with adenosine A₁ receptors. *J Pharmacol Exp Ther* 1995;273:637–642.
12. Chen C, Okayama H. High-efficiency transformation of mammalian cells by plasmid DNA. *Mol Cell Biol* 1987;7:2745–2752.
13. Chernov HI, Wilson DE, Fowler F, Plummer AJ. Non-specificity of the mouse writhing test. *Arch Int Pharmacodyn* 1967;167:171–178.
14. Clapham J, Kilpatrick GJ. Histamine H₃ receptors modulate the release of [³H]-acetylcholine from slices of rat entorhinal cortex: evidence for the possible existence of H₃ receptor subtypes. *Br J Pharmacol* 1992;107:919–923.
15. Consolo S, Arnaboldi S, Giorgi S, Russi G, Ladinsky H. 5-HT₄ receptor stimulation facilitates acetylcholine release in frontal cortex. *NeuroReport* 1994;5:1230–1232.
16. Coyle MJ. A cholinergic hypothesis for Alzheimer's disease. In: Meyer L, Nordeberg GH, eds. *Learning and Memory: Molecular Bases*. London: Pergamon Press, 1995:11–32.
17. D'Amour FE, Smith DL. A method for determining loss of pain sensation. *J Pharmacol Exp Ther* 1941;72:74–79.
18. Dei S, Bellucci C, Gualtieri F, et al. Analgesic, antimuscarinic activity and enantioselectivity of the four isomers of 3-quinclidinyl tropate as compared with the enantiomers of hyoscyamine. *Il Farmaco* 1995;50:303–309.

19. Doods H, Entzeroth M, Mayer N. Cardiosensitivity of AQ-RA 741, a novel tricyclic antimuscarinic drug. *Eur J Pharmacol* 1991;192:147-152.
20. Dörje F, Friebe T, Tacke R, Mutschler E, Lambrecht G. Novel pharmacological profile of muscarinic receptors mediating contraction of the guinea pig uterus. *Naunyn-Schmied Arch Pharmacol* 1990;342:284-289.
21. Dörje F, Wess J, Lambrecht G, Tacke R, Mutschler E, Brann MR. Antagonist binding profile of five cloned human muscarinic receptor subtypes. *J Pharmacol Exp Ther* 1991;256:727-733.
22. Eltze M, Gonne S, Riedel R, Schlotke B, Schudt C, Simon WA. Pharmacological evidence for selective inhibition of gastric acid secretion by telenzepine, a new antimuscarinic drug. *Eur J Pharmacol* 1985;112:211-224.
23. Eltze M. Muscarinic M₁ and M₂ receptors mediating opposite effects on neuromuscular transmission in rabbit vas deferens. *Eur J Pharmacol* 1988;151:205-221.
24. Eltze M, Figala V. Affinity and selectivity of biperiden enantiomers for muscarinic receptor subtypes. *Eur J Pharmacol* 1988;158:11-19.
25. Galeotti N, Ghelardini C, Gualtieri F, Dei S, Bartolini A. Effetto protettivo esercitato dal composto SM 21, liberatore di ACh, su vari modelli di amnesia. *VI National Conference of Italian Neuroscience Society*, June 1995, Milan, Italy.
26. Galeotti N, Ghelardini C, Gualtieri F, Romanelli MN, Bartolini A. Cognition enhancement by ACh releaser SM 21. *Ninth International Symposium on Cholinergic Mechanisms*, June 1995, Mainz, Germany.
27. Galeotti N, Ghelardini C, Scapecchi S, Gualtieri F, Bartolini A. Effect of SM 21 on cognitive deficits induced in the mouse by benzhexol and diazepam. *Soc Neurosci Abstr* 166.
28. Galeotti N, Ghelardini C, Bartolini A. Effect of pertussis toxin on morphine, diphenhydramine, baclofen, clomipramine and physostigmine antinociception. *Eur J Pharmacol* 1996;308:25-133.
29. Galeotti N, Ghelardini C, Bartolini A. 5-HT_{1A} agonists induce central cholinergic antinociception. *Pharmacol Biochem Behav* 1997;57:835-841.
30. George R, Haslett WL, Jendel DJ. The central action of a metabolite of tremorine. *Life Sci* 1962;1:361-363.
31. Ghelardini C, Malmberg-Aiello P, Giotti A, Malcangio M, Bartolini A. Investigation into atropine-induced antinociception. *Br J Pharmacol* 1990;101:49-54.
32. Ghelardini C, Giotti A, Malmberg-Aiello P, Bartolini A. Central cholinergic antinociception induced by the M₂ antagonist AQRA-741. *8th Camerino-Noordwijkerhout Symposium on "Trends in Receptors Research"*, September 1991. Camerino, Italy.
33. Ghelardini C, Giotti A, Gualtieri F, et al. Presynaptic auto- and hetero-receptors in the cholinergic regulation of pain. In: Angeli P, Gulini U, Quaglia W, eds. *Trends in Receptor Research*. Amsterdam: Elsevier, 1992:95-114.
34. Ghelardini C, Gualtieri F, Baldini M, et al. R-(+)-hyoscyamine: The first and selective antagonist for guinea pig uterus muscarinic receptor subtype. *Life Sci* 1993;52:569.
35. Ghelardini C, Galeotti N, Baldini M, Meoni P, Giotti A, Bartolini A. The 5-HT_{1A} agonist 8-OH-DPAT induces antinociception through a cholinergic mechanism. *Can J Physiol Pharmacol (Suppl)* 1994:380.
36. Ghelardini C, Galeotti N, Casamenti F, et al. Central cholinergic antinociception induced by 5-HT₄ agonists: BIMU 1 and BIMU 8. *Life Sci* 1996;58:2297-2309.
37. Ghelardini C, Galeotti N, Bartolini A. Caffeine induces central cholinergic analgesia. *Naunyn-Schmied Arch Pharmacol* 1997;356:590-595.
38. Ghelardini C, Galeotti N, Fantetti L, et al. Central muscarinic antinociception induced by ET-142 and SS-20 in rodents. *Drug Dev Res* 1997;42:26-34.
39. Ghelardini C, Galeotti N, Gualtieri F, et al. Antinociceptive profile of SM 21: A novel analgesic with a presynaptic cholinergic mechanism of action. *J Pharmacol Exp Ther* 1997;282:430-439.
40. Ghelardini C, Galeotti N, Gualtieri F, Romanelli MR, Bartolini A. Antinociception induced by SM 32 depends on a central cholinergic mechanism. *Pharmacol Res* 1997;35:141-147.
41. Ghelardini C, Gualtieri F, Romanelli MN, et al. Stereoselective increase in cholinergic transmission by R-(+)-hyoscyamine. *Neuropharmacology* 1997;36:281-294.
42. Giachetti A, Micheletti R, Montagna E. Cardiosensitivity profile of AF-DX 116: A muscarinic M₂ receptor antagonist. *Life Sci* 1986;38:1663-1672.
43. Giovannini MG, Casamenti F, Nistri A, Paoli F, Pepeu G. Effect of thyrotropin releasing hormone (TRH) on acetylcholine release from different brain areas investigated by microdialysis. *Br J Pharmacol* 1991;102:363-368.

44. Gorell JM, Czamecki B. Pharmacological evidence for direct dopaminergic regulation of striatal acetylcholine release. *Life Sci* 1986;38:2239–2246.
45. Gouliarov AH, Senning A. Piracetam and other structurally related nootropics. *Brain Res Rev* 1994;19:180–222.
46. Gualtieri F, Ghelardini C, Giotti A, Malcangio M, Malmberg-Aiello P, Bartolini A. Analgesia induced by the M₂ antagonist methoctramine administered i.c.v.. *Trends Pharmacol Sci Suppl* IV 1989:99.
47. Gualtieri F, Bottalico C, Calandrella A, et al. Presynaptic cholinergic modulators as potent nootropic and analgesic drugs. II. 2-phenoxy, 2-phenylthio and 2-phenylamino alkanolic acid esters. *J Med Chem* 1994;37:1712–1719.
48. Gualtieri F, Conti G, Dei S, et al. Presynaptic cholinergic modulators as potent nootropic and analgesic drugs. I. Tropic and 2-phenylpropionic acid esters. *J Med Chem* 1994;37:1704–1711.
49. Haley TJ, McCormick WG. Pharmacological effects produced by intracerebral injection of drugs in the conscious mouse. *Br J Pharmacol Chemother* 1957;12:12–15.
50. Harris LS, Dewey WL, Howes JF, Kennedy JS, Pars H. Narcotic antagonists analgesics: Interaction with cholinergic system. *J Pharmacol Exp Ther* 1969;169:17–22.
51. Hendershot LC, Forsaith J. Antagonism of the frequency of phenylquinone-induced writhing in the mouse by weak analgesics and nonanalgesics. *J Pharmacol Exp Ther* 1959;125:237–240.
52. Herz A. Wirkungen des Arecolins auf das Zentralnervensystem. *Arch Exp Path Pharmacol* 1962;242:414–429.
53. Hoehn K, Reid A, Sawynok J. Pertussis toxin inhibits antinociception produced by intrathecal injection of morphine, noradrenaline and baclofen. *Eur J Pharmacol* 1988;146:65–72.
54. Imperato A, Obinu MC, Casu A, Mascia S, Dazzi L, Gessa GL. Evidence that neuroleptics increase striatal acetylcholine release through stimulation of dopamine D₁ receptors. *J Pharmacol Exp Ther* 1993;266:557–562.
55. Jackisch R, Strittmatter H, Kasakov I, Hertting G. Endogenous adenosine as a modulator of hippocampal acetylcholine release. *Naunyn-Schmied Arch Pharmacol* 1984;327:319–325.
56. Koster R, Anderson M, De Beer EJ. Acetic acid for analgesic screening. *Fed Proc* 1959;18:412.
57. Kuribara H, Higuchi Y, Takadoro S. Effects of central depressants on rota-rod and traction performances in mice. *Jpn J Pharmacol* 1977;27:117–126.
58. Lapchak PA, Araujo DM, Quirion R, Collier B. Binding sites for [³H]AF-DX 116 and effect of AF-DX 116 on endogenous acetylcholine release from brain slices. *Brain Res* 1989;496:285–294.
59. Leighton GE, Rodriguez RE, Hill RG, Hughes J. k-Opioid agonists produce antinociception after i.v. and i.c.v. but not intrathecal administration in the rat. *Br J Pharmacol* 1988;93:553–560.
60. Lentz TL, Liley L, Michaelson U. Some actions of anticholinergic drugs. *Br J Pharmacol* 1969;32:156–162.
61. Malcangio M, Pizzighelli L, Ghelardini C, Malmberg-Aiello P, Giotti A, Bartolini A. Cross-tolerance between baclofen and bicuculline antinociception. *Pharmacol Res* 1990;22:5–6.
62. Malcangio M, Malmberg-Aiello P, Giotti A, Ghelardini C, Bartolini A. Desensitization of GABA_B receptors and antagonism by CGP 35348, prevent bicuculline- and picrotoxin-induced antinociception. *Neuropharmacology* 1992;31(8):783–791.
63. Malmberg-Aiello P, Lamberti C, Ghelardini C, Giotti A, Bartolini A. Role of histamine in rodent antinociception. *Br J Pharmacol* 1994;111:1269–1279.
64. Manetti D, Romanelli MN, Bartolini A, et al. Reduced flexibility analogues of analgesic and cognition enhancing α -tropanyl esters. *Arch Pharmacol* 1996;329:105–111.
65. McKinney M, Miller JH, Aagaard PJ. Pharmacological characterization of the rat hippocampal muscarinic autoreceptor. *J Pharmacol Exp Ther* 1993;264:74–78.
66. Melchiorre C, Cassinelli A, Quaglia W. Differential blockade of muscarinic receptor subtypes by polymethylene tetraamines. Novel class of selective antagonists of cardiac M₂ muscarinic receptors. *J Med Chem* 1987;30:201–204.
67. Meoni P, Galeotti N, Ghelardini C, et al. Protective action by the new cholinergic amplifier SM 21 on hypoxia-induced amnesia. *Workshop Meccanismi e Basi Razionali del Trattamento Farmacologico del Danno Tessutale da Ischemia-Riperfusione*, May 1993, Siena, Italy.
68. Mondadori C, Preiswerk G, Jaekel J. Treatment with a GABA_B receptor blocker improves the cognitive performance of mice, rats, and rhesus monkeys. *Pharmacol Comm* 1992;2:93–97.

69. O'Callaghan JP, Holtzman SG. Quantification of the analgesic activity of narcotic antagonists by a modified hot-plate procedure. *J Pharmacol Exp Ther* 1975;192:497-505.
70. Parenti M, Tirone F, Giagnoni G, Pecora N, Parolaro D. Pertussis toxin inhibits the antinociceptive action of morphine in the rat. *Eur J Pharmacol* 1986;124:357-359.
71. Paton WDM, Vizi ES. The inhibitory action of noradrenaline and acetylcholine output by guinea pig longitudinal muscle strip. *Br J Pharmacol* 1969;35:10-28.
72. Pedigo NW, Dewey WL, Harris LS. Determination and characterization of the antinociceptive activity of intracerebroventricularly administered acetylcholine. *J Pharmacol Exp Ther* 1975;193:845-852.
73. Prado WA, Corrado AP. Cholinergic agonist and antagonist interactions on motor nerve endings of the rat-evidence for the involvement of presynaptic receptors in the regulation of acetylcholine release. *Gen Pharmacol* 1987;18:75-81.
74. Reynell PC, Spray GH. The simultaneous measurement of absorption and transit in the gastro-intestinal tract of the rat. *J Physiol* 1956;131:452-462.
75. Richardson RT, DeLong MR. A reappraisal of the functions of the nucleus basalis of Meynert. *Trends Neurosci* 1988;11:264-267.
76. Romanelli MN, Bartolini A, Bertucci C, et al. Synthesis and enantioselectivity of the enantiomers of PG-9 and SM 21, new potent analgesic drugs. *Chirality* 1996;8:225-233.
77. Sawynok J, Reid A. Role of G-proteins and adenylate cyclase in antinociception produced by intrathecal purines. *Eur J Pharmacol* 1988;156:25-34.
78. Scatton B. Further evidence for the involvement of D₂, but not D₁ dopamine receptors in dopaminergic control of striatal cholinergic transmission. *Life Sci* 1992;31:2883-2890.
79. Stillman MJ, Shukitt-Hale B, Kong RM, Levy A, Lieberman HR. Elevation of hippocampal extracellular acetylcholine levels by methoctramine. *Brain Res Bull* 1993;32:385-89.
80. Summers RW, Kent TH, Osborne JW. Effects of drugs, ileal obstruction, and irradiation on rat gastrointestinal propulsion. *Gastroenterology* 1970;59:731-739.
81. Töröcsik A, Vizi ES. Presynaptic effects of methoctramine on release of acetylcholine. *Neuropharmacology* 1991;30:293-298.
82. Wedzony K, Limberger N, Spath L, Wichman T, Stare K. Acetylcholine release in rat nucleus accumbens is regulated through dopamine D₂-receptors. *Naunyn-Schmied Arch Pharmacol* 1988;338:250-55.
83. Wessler I, Kilbinger H. Release of [³H]acetylcholine from a modified rat phrenic nerve-hemidiaphragm preparation. *Naunyn-Schmied Arch Pharmacol* 1986;334:357-364.
84. Wessler I, Diener A, Hoffer mann M. Facilitatory and inhibitory muscarinic receptors on the rat phrenic nerve: effects of pirenzepine and dicyclomine. *Naunyn-Schmied Arch Pharmacol* 1988;338:138-142.